

### Biochemical Pharmacology

Biochemical Pharmacology 62 (2001) 1433–1438 Commentary

# Dual inhibitors of cyclooxygenase and 5-lipoxygenase. A new avenue in anti-inflammatory therapy?

Stefano Fiorucci<sup>a</sup>, Rosaria Meli<sup>b</sup>, Mariarosaria Bucci<sup>b</sup>, Giuseppe Cirino<sup>b,\*</sup>

<sup>a</sup>Sezione di Gastroenterologia ed Epatologia, Dipartimento di Medicina Clinica e Sperimentale, Università delgi Studi di Perugia, Via E.A del Pozzo, Perugia, Italy

### Abstract

Nonsteroidal anti-inflammatory drugs (NSAIDs) are a mainstay in the treatment of inflammatory disease and are among the most widely used drugs worldwide. They are anti-inflammatory, antipyretic, and analgesic and are prescribed as first choice for the treatment of rheumatic disorders and, in general, inflammation. The main limitation in using NSAIDs consists in their side-effects, including gastrointestinal ulcerogenic activity and bronchospasm. The mechanism of action of these drugs is attributed to the inhibition of cyclooxygenase (COX), and, consequently, the conversion of arachidonic acid into prostaglandins. It is hypothesized that the undesirable side-effects of NSAIDs are due to the inhibition of COX-1 (constitutive isoform), whereas the beneficial effects are related to the inhibition of COX-2 (inducible isoform). Arachidonic acid can also be converted to leukotrienes (LTs) by the action of 5-lipoxygenase (5-LOX). LTC<sub>4</sub>, LTD<sub>4</sub>, and LTE<sub>4</sub> are potent bronchoconstrictors, whereas LTB<sub>4</sub> is chemotactic for leukocytes and plays an important role in the development of gastrointestinal ulcers by contributing to the inflammatory process. Thus, developing dual inhibitor compounds that will simultaneously inhibit COX and 5-LOX could enhance their individual anti-inflammatory effects and reduce the undesirable side-effects associated with NSAIDs, especially of the gastrointestinal tract. The most promising COX/5-LOX inhibitor is ML3000 ([2,2-dimethyl-6-(4-chlorophenyl)-7-phenyl-2,3-dihydro-1H-pyrrolizine-5-yl]-acetic acid), now in Phase III clinical trials. This new approach will certainly help to unravel the mechanisms at the root of the undesirable effects of NSAIDs and to develop safer NSAIDs. © 2001 Elsevier Science Inc. All rights reserved.

Keywords: COX; 5-LOX inhibitors; Leukotrienes; COX inhibitors; 5-LOX inhibitors; Prostaglandins; NSAIDs

### 1. Introduction

NSAIDs are a mainstay in the treatment of inflammatory disease and are among the most widely used drugs world-wide [1]. They are anti-inflammatory, antipyretic, and analgesic and are prescribed as first choice in the treatment of rheumatic disorders and other degenerative inflammatory joint diseases. The common mechanism of action of this class of drugs can be attributed to the inhibition of PGG/H synthase, colloquially known as COX, and, consequently, the conversion of arachidonic acid into PGs [2]. The main limitation in using NSAIDs consists in their side-effects,

Abbreviations: NSAIDs, nonsteroidal anti-inflammatory drugs; COX, cyclooxygenase; LT, leukotriene; 5-LOX, 5-lipoxygenase; PG, prostaglandin; DFU, 5,5-dimethyl-3-(3-fluorophenyl)-4-(4-methylsuphonyl)-phenyl-2(5H)-furanone; and DFP, diisopropyl fluorophosphate.

including gastrointestinal ulcerogenic activity and bronchospasm [3,4]. It is now well established that COX exists as two isoforms in mammals, a constitutive form (COX-1), and an inducible form (COX-2). COX-1 is expressed in most mammalian cells under physiological conditions [5], and particularly large amounts are produced by endothelial cells, platelets, and kidney tubule cells. COX-2 is induced by pro-inflammatory stimuli such as cytokines, bacterial lipopolysaccharide, growth factors, and tumour-promoting agents [6,7]. However, COX-2 also has been shown to be constitutively expressed in the cells of the reproductive tract and in the nervous system [8]. It is currently hypothesized that the undesirable side-effects of NSAIDs are due to COX-1 inhibition, whereas the beneficial effects, such as the reduction of swelling and analgesia, are related to COX-2 inhibition [9,10]. Nevertheless, both COX-1 and COX-2 have the same substrate, i.e. arachidonic acid, from which are formed not only PGs but also LTs. The LTs are formed from arachidonic acid by the action of 5-LOX.

<sup>&</sup>lt;sup>b</sup>Dipartimento di Farmacologia Sperimentale, Università delgi Studi di Napoli-Federico II, Via Domenico Montesano 49, 80131 Napoli, Italy

<sup>\*</sup> Corresponding author. Tel.: +39 081-678442; fax: +39 081-678403 *E-mail address:* cirino@unina.it (G. Cirino).

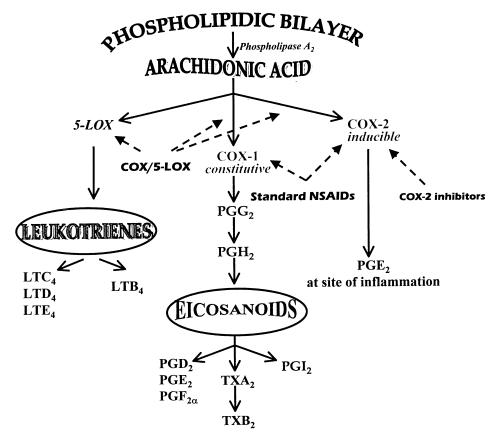


Fig. 1. Schematic pathways of the arachidonic cascade. Solid lines represent metabolite production; dotted lines represent inhibition.

While LTC<sub>4</sub>, LTD<sub>4</sub>, and LTE<sub>4</sub> are potent bronchoconstrictors [11], LTB<sub>4</sub> is a potent chemotactic agent for leukocytes, and it has been reported to play an important pathophysiological role in the development of gastrointestinal ulcers [12]. The current therapeutic approach and chemical design of NSAIDs are targeted to developing selective COX inhibitors (Fig. 1). However, it is intuitive that LTs, and in particular LTB<sub>4</sub>, which is involved in leukocyte recruitment at the site of injury, also contribute and sustain the inflammatory process at the site of the injury. For this reason, developing compounds that will inhibit COX and 5-LOX at the same time could lead to an enhanced anti-inflammatory effect and reduce undesirable side-effects (Fig. 1).

## 2. COX-1 and COX-2 inhibition: a double-edged sword

One of the most important inflammatory diseases associated with COX-2 induction is arthritis. Indeed, in animal models of arthritis, COX-2 is strongly expressed, and it seems to be responsible for the increase in PG production [13]. COX-2 expression has also been found in human osteoarthritis [14], as well as in synovial tissue obtained from patients with rheumatoid arthritis [15]. In addition, several studies have also suggested a potential role for

COX-2 in inflammatory pain, since highly selective COX-2 inhibitors (e.g. DFU) inhibit hyperalgesia in rats [16], and pain-inducing sensory stimulation leads to induction of COX-2 in the spinal cord, as shown in rats exposed to inflammatory stimuli [17,18]. In humans, selective COX-2 inhibitors, such as rofecoxib, have been shown to be analgesic in a post-dental surgery pain study [19]. All of these studies indicate selective COX-2 inhibitors as drugs that might, in theory, reduce the severity of adverse reactions of non-selective NSAIDs. However, recent studies have shown that the role played by COX-1 is more than physiological, just as the role played by COX-2 is not exclusively pathological. Recently, Warner and coworkers [20] using different in vitro assays investigated the relative potencies, as inhibitors of COX-1 and COX-2, of a wide range of NSAIDs as well as the newer COX-2 selective agents. The authors, on the basis of the results obtained, classified the agents tested into four main groups, which range from compounds that are full inhibitors of both COX-1 and COX-2 (group one) to compounds that selectively inhibit COX-2 with only weak activity against COX-1 (group four) [20]. Comparing these groups of NSAIDs to the epidemiological studies of NSAID-induced gastrointestinal toxicity [21], it has been found that compounds associated with the greatest gastrointestinal toxicity have the greatest COX-1 selectivity. It is of particular interest to note that this group includes indomethacin, ketoprofen, naproxen, and ketorolac, which are among the most efficacious NSAIDs used as pain killers. In this respect, a recent paper has shed new light on the role played by COX-1 in pain perception, and it has confirmed this previous observation. By using knockout mice homozygous for COX-1<sup>-/-</sup> and for COX-2<sup>-/-</sup>, it has been proposed that peripheral COX-1 mediates nociception in slowly developing pain, while spinal COX-1 is involved in rapidly transmitting pain [22]. However, compounds that selectively inhibit COX-2 (>50-fold COX-2 selective), such as rofecoxib and DFP, have been shown to be effective as anti-inflammatory drugs and to produce few serious gastrointestinal complications when used in the general population; preliminary reports indicate that rofecoxib has low gastrointestinal toxicity, but clinical trials are still in progress to confirm this finding [23]. Nevertheless, also concerning gastric safety, it has to be taken into account that studies in animals [24] and in humans [25] have shown that COX-2 selective inhibitors can delay the repair of existing gastrointestinal damage caused by ulcers, most likely by reducing the synthesis of PGs, produced by COX-2, that are thought to play an important role in the healing process. Thus, chronic use of COX-2 selective inhibitors may delay the healing of pre-existing ulcers. Another beneficial effect proposed for COX-2 is the protective role that this enzyme may play in asthma; in fact, it has been demonstrated that COX-2 is involved in controlling human airway smooth muscle cell proliferation [26]. In addition, it has also been shown that COX-2 is expressed in the airways of aspirinsensitive patients [27,28], implying that COX-2 induction in airway cells in some way may limit the production of LTs [25]. Indeed, it is well known that aspirin and other NSAIDs can induce asthmatic symptoms in sensitive patients [29].

Thus, it appears that selective COX-2 inhibitors do not represent a complete answer to the need for safer and more effective drugs to be used in inflammatory disease therapy.

### 3. Role of 5-LOX

The activation of phospholipase A<sub>2</sub> induces the mobilization of fatty acids from the membrane lipid pool, in particular arachidonic acid, for the synthesis of lipidic mediators at the site of cellular damage. As discussed above, arachidonic acid is metabolized by two major pathways to pro-inflammatory mediators: the COX pathway and the 5-LOX pathway [30,31]. The first step in the 5-LOX cascade consists of activation of the enzyme by 5-LOX-activating protein (FLAP), which leads to the formation of the LTs [32] 5-hydroperoxyeicosatetraenoic acid (5-HPETE) and 5-hydroxyeicosatetraenoic acid (5-HETE). LTC<sub>4</sub>, LTD<sub>4</sub>, and LTE<sub>4</sub> are known as "cysteinyl leukotrienes" because of the presence of a thioether-linked peptide, and there is some evidence supporting the role of sulphidopeptide-LT in the pathophysiology of asthma [33]. These LTs also induce the synthesis and release of other pro-inflammatory mediators such as interleukin (IL)-8 and plateletactivating factor [34]. LTB<sub>4</sub> has remarkable chemotactic activity on neutrophils [35] and eosinophils [36,37], and it also has been shown to be involved in the pathogenesis of a wide variety of human inflammatory diseases [38,39]. The role played by LTs in these pathologies has been addressed in recent years by using LT receptor antagonists [40,41]. Indeed, LT receptor antagonists have been shown to be effective in the maintenance treatment of patients with mildto-moderate persistent asthma [34]. Evidence has accumulated in recent years for a key role for LTB4 in the development of the inflammatory response. Recently, Haribabu and coworkers [42] demonstrated that deletion of the LTB<sub>4</sub> receptor in mice leads to a reduction in the vascular and cellular components of acute inflammatory responses, suggesting inhibition of LTB<sub>4</sub> activity as a therapeutic intervention in certain human inflammatory conditions [42]. In addition, the recent discovery of a new distinct receptor for LTB<sub>4</sub> in humans and mice may lead to the identification of other members of the LTB<sub>4</sub> receptor family and provide new knowledge concerning the pathophysiological role of LTB<sub>4</sub> [43].

### 4. Dual inhibitors of COX/5-LOX

Inhibition of 5-LOX is considered the ideal treatment for allergic disorders and asthma [44]. Indeed, it is known that COX inhibitors induce adverse reactions in patients with asthma either because of the shunting of LT synthesis due to increased availability of arachidonic acid to the 5-LOX pathway [44-46] or because of a reduction in vasodilatory prostaglandins PGE<sub>2</sub> and PGI<sub>2</sub> that have been shown to protect the airways [47–49]. A promising alternative to avoiding the adverse reactions of NSAIDs could be the development of drugs that inhibit both COX and 5-LOX (COX/5-LOX), leading to compounds with improved efficacy and reduced side-effects when compared with selective COX inhibitors [50]. In addition, dual inhibitors of the COX/5-LOX pathways may exhibit anti-inflammatory activity with a wider spectrum than that of classical NSAIDs by inhibiting 5-LOX product-mediated inflammatory reactions towards which NSAIDs are ineffective [51,52]. To this end, Inagaki et al. [53] recently found a novel antiarthritic agent, S-2474 (Fig. 2). This compound displayed a dual inhibition of COX/5-LOX with good selectivity toward COX-2 inhibition, like celecoxib. In rats, it exerted excellent anti-inflammatory activity without ulcerogenic effects and showed cytokine-modulating properties in THP-1 cells [53].

ER-34122, 5-{[1,5-bis(4-methoxyphenyl)pyrazol-3-yl] dimethoxymethyl}-2-chlorobenzamide (Fig. 2), is an orally active dual inhibitor of COX/5-LOX, and *in vivo* in mice this molecule has shown an enhanced anti-inflammatory effect, when compared with indomethacin, that has been linked to its ability to inhibit the generation of 5-LOX

Fig. 2. Molecular structures of dual inhibitors

products [52]. These promising data have been further confirmed by the finding that ER-34122 suppresses polymorphonuclear neutrophil (PMN) infiltration, subsynovial soft tissue oedema, and multiplication of synovial lining cells in the early stage of arthritis in MRL/l mice [51]. Tepoxalin (Fig. 2), a dual inhibitor compound discontinued in clinical Phase II, significantly inhibited gastric LTB<sub>4</sub> synthesis in rats at a dose of 10 mg/kg, and at the same dose it markedly suppressed PG synthesis at a site of peripheral inflammation [54]. Tepoxalin also inhibited IL-2, IL-6, and tumor necrosis factor- $\alpha$  (TNF $\alpha$ ) production with an IC<sub>50</sub> of 10–12 mM [55]. Given orally, tepoxalin exhibited anti-inflammatory activity in rats with adjuvant-induced arthritis ( $ED_{50} = 3.5$ mg/kg) [56]. This effect was also accompanied by potent analgesic activity in an acetic acid-induced abdominal constriction assay in the mouse ( $ED_{50} = 0.45 \text{ mg/kg}$ ) [56].

Among dual inhibitors, a novel class of non-antioxidant compounds has been described; the most potent and wellbalanced dual inhibitor of COX/5-LOX is ML3000 ([2,2dimethyl-6-(4-chlorophenyl)-7-phenyl-2,3-dihydro-1H-pyrrolizine-5-yl]-acetic acid; see Fig. 2), a derivative of the phenyl moiety at the 6-position of the pyrrolizine ring [57], now in clinical Phase III. The inhibition of COX was determined in a bovine thrombocyte intact cell assay and inhibition of 5-LOX was assayed in intact bovine polymorphonuclear leukocytes ( $IC_{50} = 0.21 \mu M$ , COX; 0.18  $\mu M$ , 5-LOX). ML3000 and indomethacin have been compared in a number of experimental models of inflammation. In carrageenan-induced rat paw oedema, the ED<sub>50</sub> of ML3000 for oral administration was 17 mg/kg versus 3 mg/kg for indomethacin [58]. While indomethacin was more potent in this model, ML3000 produced significantly less gastric injury.

ML3000 significantly inhibited whole blood thromboxane synthesis and, at doses which have anti-inflammatory activity, significantly suppressed gastric PG synthesis without producing significant mucosal injury [59]. In a guinea pig model of arachidonic acid bronchoconstriction, ML3000 was found to be highly effective and potent ( $ED_{50} = 0.2$ mg/kg) [60]; when administered as an aerosol at a dose of 100 mg, 30 min before antigen challenge in allergic sheep, ML3000 provided significant inhibition against the early bronchial response, completely blocking the late antigeninduced bronchoconstriction and the airway responsiveness to aerosolized carbachol that occur 24 hr after antigen challenge in this model [49]. In the rat, oral administration of ML3000 at doses of 30-300 mg/kg produced no gastrointestinal damage [61]. After 11 days of daily dosing, some rats treated with ML3000 showed duodenal damage, but the level of damage caused by indomethacin was more severe and occurred at lower doses [60]. Thus, while clinical data are needed to confirm the same profile in humans, animal data suggest that COX/5-LOX inhibitors may represent a valid therapeutic alternative to standard NSAIDs and selective COX-2 inhibitors.

### 5. Concluding remarks

It is beyond question that in the NSAID field there is no need to develop more powerful drugs since they are already available. Indeed, the therapeutic demand has been always directed towards safer NSAIDs. The discovery of the inducible form of COX, i.e. COX-2, has given an enormous boost to research in this field, and COX-2 inhibitors have

been developed to fill the gap between therapeutic demand and the NSAIDs available in therapy. Thus, the concept of COX-2 selective inhibition being the key to new effective but safer NSAIDs has led to the development of very selective inhibitors that are now available on the market. The "drum beat" of companies that have developed these compounds has created the general feeling that the therapeutic problem linked to the side-effects of NSAIDs, which represent the major limitation to their use, is on its way to being solved. However, as often happens in science, things are never as simple as they appear. Indeed, a physiological role for COX-2 has been proposed, and a key role in pain perception for COX-1 has been put forward recently. Thus, COX-1 is not playing just a physiological role and COX-2 is not playing solely a pathological role, and this Ying-Yang theory has to be revised carefully. Inflammation is a very complex phenomenon, and several mediators play an important role in its development at the site of the injury. Currently, there are other approaches to this problem that are being developed to obtain NSAIDs that are effective but devoid of their major adverse side-effects. In this regard, the development of COX/5-LOX inhibitors may represent a new promising alternative. Indeed, by inhibiting both pathways these compounds may lead to a better anti-inflammatory effect accompanied by reduced gastric side-effects. In addition, since a key role in inflammation is also played by oxygen radicals, it must be taken into consideration that inhibition of the COX pathway increases their formation through the peroxidative cleavage of 5-HETEs and that 5-LOX inhibition would attenuate this effect. Certainly the development of these inhibitors, as well as of other different approaches, will help to unravel the complex mechanisms at the root of the undesirable effects of NSAIDs and aid in developing newer and safer NSAIDs.

### References

- Garner A. Adaptation in the pharmaceutical industry, with particular reference to gastrointestinal drugs and diseases. Scand J Gastroenterol 1992;27(Suppl 193):83.
- [2] Vane JR. Inhibition of prostaglandin synthesis as a mechanism of action for aspirin-like drugs. Nat New Biol 1971;231:232–9.
- [3] Amadio P, Cummings DM, Amadio P. Nonsteroidal anti-inflammatory drugs. Tailoring therapy to achieve results and avoid toxicity. Postgrad Med 1993;93:73–97.
- [4] Wallace JL, Carter E, McKnight GW, Le T, McCafferty DM, Argentieri D, Capetola R. Tissue-selective inhibition of prostaglandin (PG) synthesis: anti-inflammatory without gastropathy? Gastroenterology 1993;104:A221.
- [5] Seibert K, Zhang Y, Leahy K, Hauser S, Masferrer J, Isakson P. Distribution of COX-1 and COX-2 in normal and inflammed tissues. Adv Exp Med Biol 1997;400A:167–70.
- [6] Vane JR, Bakhle YS, Botting RM. Cyclooxygenases 1 and 2. Annu Rev Pharmacol Toxicol 1998;38:97–120.
- [7] Fu J-Y, Masferrer JL, Seibert K, Raz A, Needleman P. The induction and suppression of prostaglandin H<sub>2</sub> synthase (cyclooxygenase) in human monocytes. J Biol Chem 1990;265:16737–40.
- [8] Yamagata K, Andreasson KI, Kaufmann WE, Barnes CA, Worley PF. Expression of a mitogen-inducible cyclooxygenase in brain neurons:

- regulation by synaptic activity and glucocorticoids. Neuron 1993;11: 371–86.
- [9] Mitchell JA, Akarasereenont P, Thiemermann C, Flower RJ, Vane JR. Selectivity of nonsteroidal antiinflammatory drugs as inhibitors of constitutive and inducible cyclooxygenase. Proc Natl Acad Sci USA 1993;90:11693–7.
- [10] Smith WL, Garavito M, DeWitt DL. Prostaglandin endoperoxide H synthases (cyclooxygenases)-1 and -2. J Biol Chem 1996;271:33157– 60
- [11] Bisgaard H. Leukotrienes and prostaglandins in asthma. Allergy 1984;39:413–20.
- [12] Asako H, Kubes P, Wallace J, Gaginella T, Wolf RE, Granger DN. Indomethacin-induced leukocyte adhesion in mesenteric venules: role of lipoxygenase products. Am J Physiol 1992;262:G903–8.
- [13] Anderson GD, Hauser SD, McGarity KL, Bremer ME, Isakson PC, Gregory SA. Selective inhibition of cyclooxygenase (COX)-2 reverses inflammation and expression of COX-2 and interleukin-6 in rat adjuvant arthritis. J Clin Invest 1996;97:2672–9.
- [14] Amin AR, Attur M, Patel RN, Thakker GD, Marshall PJ, Rediske J, Stuchin SA, Patel IR, Abramson SB. Superinduction of cyclooxygenase-2 activity in human osteoarthritis-affected cartilage. Influence of nitric oxide. J Clin Invest 1997;99:1231–7.
- [15] Kang RY, Freire-Moar J, Sigal E, Chu CQ. Expression of cyclooxygenase-2 in human and an animal model of rheumatoid arthritis. Br J Rheumatol 1996;35:711–8.
- [16] Riendeau D, Percival MD, Boyce S, Brideau C, Charleson S, Cromlish W, Ethier D, Evans J, Falgueyret JP, Ford-Hutchinson AW, Gordon R, Greig G, Gresser M, Guay J, Kargman S, Leger S, Mancini JA, O'Neill G, Ouellet M, Rodger IW, Therien M, Wang Z, Webb JK, Wong E, Chan CC. Biochemical and pharmacological profile of a tetrasubstituted furanone as a highly selective COX-2 inhibitor. Br J Pharmacol 1997;121:105–17.
- [17] Beiche F, Scheurer S, Brune K, Geisslinger G, Goppelt-Struebe M. Up-regulation of cyclooxygenase-2 mRNA in the rat spinal cord following peripheral inflammation. FEBS Lett 1996;390:165–9.
- [18] Gardiner NJ, Gilett S, Grubb BD. Cyclooxygenase in rat spinal cord: selective induction of COX-2 during peripheral inflammation. Br J Pharmacol 1999;120:71P.
- [19] Morrison BW, Christensen S, Yuan W, Brown J, Amlani S, Seidenberg B. Analgesic efficacy of the cyclooxygenase-2-specific inhibitor rofecoxib in post-dental surgery pain: a randomized controlled trial. Clin Ther 1999;21:943–53.
- [20] Warner TD, Giuliano F, Vojnovic I, Bukasa A, Mitchell JA, Vane JR. Nonsteroid drug selectivities for cyclo-oxygenase-1 rather than cyclo-oxygenase-2 are associated with human gastrointestinal toxicity: a full in vitro analysis. Proc Natl Acad Sci USA 1999;96:7563-8.
- [21] Henry D, Lim LL-Y, Garcia Rodriguez LA, Perez Gutthann S, Carson JL, Griffin M, Savage R, Logan R, Moride Y, Hawkey C, Hill S, Fries JT. Variability in risk of gastrointestinal complications with individual non-steroidal anti-inflammatory drugs: results of a collaborative meta-analysis. BMJ 1996;312:1563–6.
- [22] Ballou LR, Botting RM, Goorha S, Zhang J, Vane JR. Nociception in cyclooxygenase isoenzyme-deficient mice. Proc Natl Acad Sci USA 2000;97:10272–6.
- [23] Hawkey CJ. COX-2 inhibitors. Lancet 1999;353:307-14.
- [24] Reuter BK, Asfaha S, Buret A, Sharkey KA, Wallace JL. Exacerbation of inflammation-associated colonic injury in rat through inhibition of cyclooxygenase-2. J Clin Invest 1996;98:2076–85.
- [25] Mitchell JA, Warner TD. Cyclooxygenase-2: pharmacology, physiology, biochemistry and relevance to NSAID therapy. Br J Pharmacol 1999;128:1121–32.
- [26] Belvisi MG, Saunders M, Yacoub M, Mitchell JA. Cyclooxygenase-2 induction has a profound inhibitory effect on human airway smooth muscle cell proliferation. Br J Pharmacol 1998;125:1102–8.
- [27] Sousa AR, Pfister R, Christie PE, Lane SJ, Nasser SM, Schmitz-Shumann M, Lee TH. Enhanced expression of cyclooxygenase isoen-

- zyme 2 (COX-2) in asthmatic airways and its cellular distribution in aspirin-sensitive asthma. Thorax 1997;52:940-5.
- [28] Cowburn AS, Sladek K, Soja J, Adamek L, Nizankowska E, Szczeklik A, Lam BK, Penrose JF, Austen FK, Holgate ST, Sampson AP. Overexpression of leukotriene C<sub>4</sub> synthase in bronchial biopsies from patients with aspirin-intolerant asthma. J Clin Invest 1998;101:834– 46.
- [29] Kowalski ML. Aspirin sensitive rhinosinusitis and asthma. Allergy Proc 1995;16:77–80.
- [30] Weber PC. Fish oil fatty acid and cardiovascular function: epidemiology and biochemical mechanisms. Biochem Soc Trans 1993;18: 1045–9
- [31] Neuhof H, Seeger W, Suttorp N. Activation of the pulmonary arachidonic acid system and its consequences for hemodynamics and fluid balance. In: Schalag G, Redl H, editors. Pathophysiological role of mediators and mediator inhibitors in shock (First Vienna Shock Forum). New York: Alan R. Liss, 1987. p. 289–300.
- [32] Lewis RA, Austen KF, Drazen JM, Clark DA, Marfat A, Corey EJ. Slow-reacting substances of anaphylaxis: identification of leukotrienes C-1 and D from human and rat sources. Proc Natl Acad Sci USA 1980;77:3710-4.
- [33] Chanarin N, Johnston SL. Leukotrienes as a target in asthma therapy. Drugs 1994;47:12–24.
- [34] Papi A, Caramori G, Fabbri LM. Current asthma therapies and issues in asthma management. Eur Respir Rev 1998;8:341–7.
- [35] Ford-Hutchinson AW, Bray MA, Doig MV, Shipley ME, Smith MJH. Leukotriene B, a potent chemokinetic and aggregating substance released from polymorphonuclear leukocytes. Nature 1980;286:264-5.
- [36] Huang W-W, Garcia-Zepeda EA, Sauty A, Oettgen ME, Rothenberg HC, Luster AD. Molecular and biological characterization of murine leukotriene B<sub>4</sub> receptor expressed on eosinophils. J Exp Med 1998; 188:1063–74.
- [37] Ng CF, Sun FF, Taylor BM, Wolin MS, Wong PYK. Functional properties of guinea pig eosinophil leukotriene B<sub>4</sub> receptor. J Immunol 1991;147:3096–103.
- [38] Samuelsson B, Dahlen SE, Lindgren JA, Rouzer CA, Serhan CN. Leukotrienes and lipoxins: structures, biosynthesis, and biological effects. Science 1987;237:1171–6.
- [39] Lewis RA, Austen KF, Soberman RJ. Leukotrienes and other products of the 5-lipoxygenase pathway. Biochemistry and relation to pathobiology in human diseases. N Engl J Med 1990;323:645–55.
- [40] Tager AM, Dufour JH, Goodarzi K, Bercury SD, von Andrian UH, Luster AD. BLTR mediates leukotriene B<sub>4</sub>-induced chemotaxis and adhesion and plays a dominant role in eosinophil accumulation in a murine model of peritonitis. J Exp Med 2000;192:439-46.
- [41] Devchand PR, Kekker H, Peters JM, Vazquez M, Gonzalez FJ, Wahli W. The PPARα-leukotriene B<sub>4</sub> pathway to inflammation control. Nature 1996;384:39-43.
- [42] Haribabu B, Verghese MW, Steeber DA, Sellars DD, Bock CB, Snyderman R. Targeted disruption of the leukotriene B<sub>4</sub> receptor in mice reveals its role in inflammation and platelet-activating factorinduced anaphylaxis. J Exp Med 2000;192:433–8.
- [43] Yokomizo T, Kato K, Terawaki K, Izumi T, Shimizu T. A second leukotriene B<sub>4</sub> receptor, BLT<sub>2</sub>. A new therapeutic target in inflammation and immunological disorders. J Exp Med 2000;192:421–31.
- [44] Rabasseda X, Mealy N, Castañer J. ML3000. Drugs of the future 1995;20:1007–9.
- [45] Kuehl FA, Daugherty HW, Ham EA. Interaction between prostaglandins and leukotrienes. Biochem Pharmacol 1984;33:1–5.
- [46] Dworski R, Sheller JR, Wickersham NE, Oates JA, Brigham KL, Roberts LJ, Fitzgerald GA. Allergen-stimulated release of mediators

- into sheep bronchoalveolar lavage fluid. Effect of cyclooxygenase inhibition. Am Rev Respir Dis 1989;139:46–51.
- [47] Birnbaum JE, Birkhead NC, Oronsky AL, Dessy F, Rihoux JP, VanHumbeeck L. Bronchodilator activity of a PGE<sub>2</sub> analog in animals and in man. Prostaglandins 1981;21:457–69.
- [48] Szczeklik A, Mastalerz L, Nizankowska E, Cmiel A. Protective and bronchodilator effects of prostaglandin E and salbutamol in aspirininduced asthma. Am J Respir Crit Care Med 1996;153:567–71.
- [49] Abraham WM, Laufer S, Tries S. The effects of ML3000 on antigeninduced responses in sheep. Pulm Pharmacol Ther 1997;10:167–73.
- [50] Carty TJ, Marfat A, Rasamune J. Modulation of AA metabolites in the treatment of rheumatoid arthritis. In: Allen RC, editor. Annual reports in medicinal chemistry. New York: Academic Press, 1988. p. 181–9.
- [51] Horizoe T, Nagakura N, Chiba K, Shirota H, Shinoda M, Numata H, Kobayashi S, Abe C. Effect of ER-34122, a novel dual 5-lipoxygenase/cyclooxygenase inhibitor, on indices of early articular lesion in MRL/MpJ-lpr/lpr mice. Inflamm Res 1999;48:432–6.
- [52] Horizoe T, Nagakura N, Chiba K, Shirota H, Shinoda M, Kobayashi N, Numata H, Okamoto Y, Kobayashi S. ER-34122, a novel dual 5-lipoxygenase/cyclooxygenase inhibitor with potent anti-inflammatory activity in an arachidonic acid-induced ear inflammation model. Inflamm Res 1998;47:375–83.
- [53] Inagaki M, Tsuri T, Jyoyama H, Ono T, Yamada K, Kobayashi M, Hori Y, Arimura A, Yasui K, Ohno S, Kakudo S, Koizumi K, Suzuki R, Kawai S, Kato M, Matsumoto S. Novel antiarthritic agents with 1,2-isothiazolidine-1,1-dioxide (γ-sultam) skeleton: cytokine suppressive dual inhibitors of cyclooxygenase-2 and 5-lipoxygenase. J Med Chem 2000;43:2040 – 8.
- [54] Wallace JL, McCafferty DM, Carter L, McKnight W, Argentieri D. Tissue-selective inhibition of prostaglandin synthesis in rat by tepoxalin: antiinflammatory without gastropathy? Gastroenterology 1993;105:1630-6.
- [55] Ritchie DM, Argentieri DC, Aparicio BL, Plante RK, Lau CY, Barbone AG. Cytokine-modulating activity of tepoxalin, a new potential antirheumatic. Int J Immunopharmacol 1995;17:805–12.
- [56] Argentieri DC, Ritchie DM, Ferro MP, Kirchner T, Wachter MP, Anderson DW, Rosenthale ME, Capetola RJ. Tepoxalin: a dual cycloxygenase/5-lipoxygenase inhibitor of arachidonic acid metabolism with potent anti-inflammatory activity and a favorable gastrointestinal profile. J Pharmacol Exp Ther 1994;271:1399–408.
- [57] Laufer SA, Augustin J, Dannhardt G, Kiefer W. (6,7-Diaryldihydro-pyrrolizin-5-yl)acetic acids, a novel class of potent dual inhibitors of both cyclooxygenase and 5-lipoxygenase. J Med Chem 1994;37: 1894-7
- [58] Laufer S, Tries S, Augustin J, Elsasser R, Albrecht W, Guserle R, Algate DR, Atterson PR, Munt PL. Acute and chronic anti-inflammatory properties of [2,2-dimethyl-6-(4-chlorophenyl)-7-phenyl-2,3-dihydro-1H-pyrrolizine-5-yl]-acetic acid. Arzneimittelforschung 1995;45:27–32.
- [59] Wallace JL, Carter L, McKnight W, Tries S, Laufer S. ML 3000 reduces gastric prostaglandin synthesis without causing mucosal injury. Eur J Pharmacol 1994;271:525–31.
- [60] Laufer S, Tries S, Augustin J, Elsasser R, Algate DR, Atterson PR, Munt PL. Gastrointestinal tolerance of 2,2-dimethyl-6-(4-chlorophenyl)-7-phenyl-2,3-dihydro-1H-pyrrolizine-5-yl]-acetic acid in rat. Arzneimittelforschung 1994;44:1329–33.
- [61] Algate DR, Augustin J, Atterson PR, Beard DJ, Jobling CM, Laufer S, Munt PL, Tries S. General pharmacology of [2,2-dimethyl-6-(4-chlorophenyl)-7-phenyl-2,3-dihydro-1H-pyrrolizine-5-yl]-acetic acid in experimental animals. Arzneimittelforschung 1995;45:159-65.